

Mechanisms of Image Processing in the Visual Cortex

C. ELIZABETH BOUDREAU AND DAVID FERSTER

ABSTRACT Orientation selectivity in primary visual cortex represents a crucial step in the transformation of patterns of light incident on the retina into features that define recognizable objects in the visual world. Orientation-selective responses in cortical cells can be explained well by the pattern of inputs onto the cells, indicating that the design of the network, rather than highly specialized computational abilities of individual cells, may be responsible for creating and refining their very varied and precise response properties. In addition, most properties of cells in primary visual cortex can be accounted for by a model that is largely linear and processes information primarily in one direction. These findings also indicate that the elaboration of complex feature selectivity in the visual cortex results from the careful organization of simple mathematical units.

If the visual cortex dramatically transforms the neuronal representation of the visual image, the neurons of the retina and lateral geniculate nucleus (LGN) encode the contrast of the image at each point, more or less independently of the size of the image elements that give rise to local contrast. Yet the neurons of the primary visual cortex (V1), which receive the bulk of their information about the visual image directly from the LGN, encode information about the size, direction, and depth of image features that fall within their receptive fields. None of this information is contained explicitly in the activity of single geniculate cells, so the cortical neurons are clearly integrating information they receive, either directly or indirectly, in a large number of geniculate inputs. In the cat, this information emerges almost fully articulated at the first cortical synapse, between the axons of geniculate relay neurons and the cortical recipient cells in layer 4. A detailed description of the cortical transformation raises questions about its purpose. How does it benefit an organism to parse the visual image into elements of different information? What new abilities or limitations in visual functions arise from such an organization? But the visual information that occurs in V1 is also a compelling model of cortical information processing. The image transformation that occurs there is at once complex, and yet mathematically tractable and easily described, unlike the more

ALBERT BODREAU and DAVID FERSTER Department of Neurology, Northwestern University, Evanston, Ill.

mysterious processes that underlie higher cognitive tasks such as object recognition, speech, or intelligence. For those interested in how the cortex performs neuronal computations, then, V1, and in particular the way in which cortical circuitry gives rise to orientation selectivity, has been of intense and prolonged interest. The 40 years following Hubel and Wiesel's first description of cortical receptive fields have seen a remarkable and productive interaction between theory and experiment. The process has refined our understanding of the function of a fascinating and complex piece of neuronal circuitry.

Most cells in V1 are orientation selective, meaning that they are most active (produce the most action potentials) in response to the presentation of a contrast edge at a particular orientation (e.g., vertical), while edges of other orientations produce a smaller response. The relationship between response size and stimulus orientation for individual cells can be well described by Gaussian or bell-shaped mathematical functions. In cat V1, tilting a stimulus 10° away from the optimal orientation reduces a cell's response by 50% on average. As Hubel and Wiesel first demonstrated, cells within a single vertical column of the cortex all have the same preferred orientation, and the preferred orientation of each point in cortex forms an orderly map. Although orientation selectivity is a property common to the primary visual cortices of many animal species, it is not always implemented in exactly the same way. In the cat, orientation selectivity is prevalent in all layers of primary visual cortex, and the narrowness of this selectivity does not change significantly as a function of laminar position. In the monkey, orientation selectivity is weakest in layer 4CB, where axons of LGN neurons terminate (Callaway, 1998; but see Ringach et al., 2002). In the ferret, as in the cat, cells receiving direct input from the LGN are very selective for orientation, on average (Ts'o et al., 2003), while in the tree shrew orientation selectivity is largely absent in layer 4, emerging more strongly in layers to which layer 4 neurons project (Chisum et al., 2003). Whether these interspecies differences point to substantial variation in the mechanisms for producing orientation-selective responses is not known.

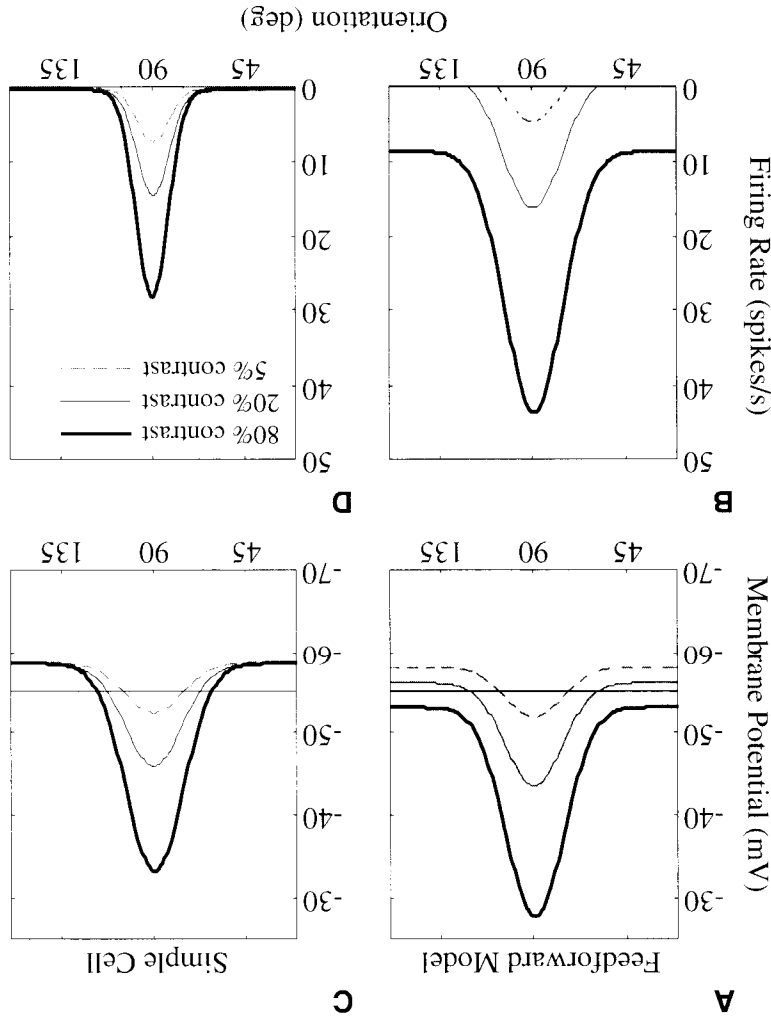
LGN relay cells exhibit weak or no selectivity for orientation, responding optimally instead to a light (ON-center

proportionally, so that they continue to balance each other and result in no net excitation of the cortical cell. Because there is no physiological evidence for direct inhibition from the LGN to VI (Freund et al., 1989; Callaway, 1998), antiphase inhibition would be mediated by inhibitory connections between cortical cells with complementary receptive field structures. Push-pull inhibition in response to the preferred stimulus orientation is well demonstrated by intracellular recordings of membrane potential and conductance (Fester, 1988; Hirsch et al., 1998). In these studies, receptive fields were mapped with light and dark spots, and then responses to stimuli of the nonpreferred polarity were tested at different locations within the receptive field. The results showed that, when a light stimulus was placed in an OFF subregion, for

contrast increases, since the activity in geniculate relay cells also The amplitude of the curves grows equally at all orientations as cells arranged in a simple cells by synaptic input from geniculate relay tuning. (d) The orientation tuning curves of membrane potential

example, the total synaptic input to the cortical cell increased and the cell membrane hyperpolarized, implying that the increase was due to greater inhibitory input. Although a reduction in excitation would also cause a hyperpolarization of the membrane, this would be associated with a decrease, not an increase, in total synaptic input. Evidence for strong or dominant inhibition at the null orientation is more equivocal. Monier and colleagues (2003) see strong cross-oriented inhibition in a subset of cortical cells. Fester (1986) and Anderson, Carandini, and colleagues (2000) found that inhibitory and excitatory input to cortical cells are tuned to the same orientation and have similar orientation tuning width. Although there was some inhibition evoked at nonpreferred orientations, it is not clear that the observed inhibition was strong enough to explain

increases with increasing stimulus contrast. (b) The orientation tuning of the cell's spike output at different contrast, as predicted from applying a fixed threshold to the tuning curves in A. (c and d) A schematic of the orientation tuning recorded in cortical simple cells (C, membrane potential; D, spike rate).

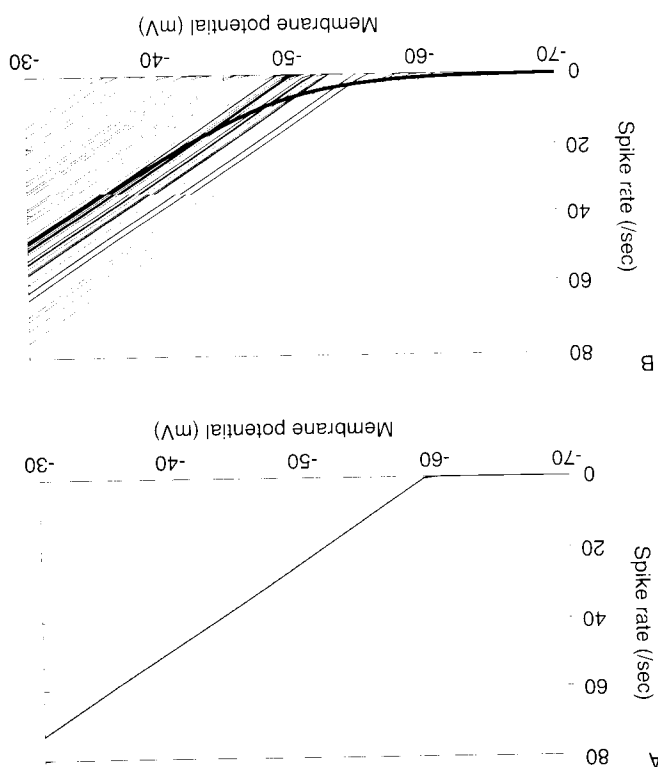


Another explanation for the generation of orientation selectivity relies heavily on the organization, both laminar and columnar, of cortex. This class of models is known as feedback models, and their orientation tuning derives from recurrent excitatory connections that amplify responses to the preferred orientation and lateral interactions that damp down unwanted responses to the null orientation (Ben-Yishai, Bar-On, and Sompolinsky, 1995; Somers, Nelson, and Sur, 1995). A generalized schematic of feedback model connectivity is shown in figure 21.4.

Feedback models have the advantage of requiring very simple rules for the connections between neurons. They can be implemented in networks in which cells are excited by and inhibited by all of their neighbors, with the strength of these connections declining uniformly with increasing distance between cells (McLaughlin et al., 2000). Others propose excitatory connections between cells of like

Feedback models of orientation selectivity

Figure 21.3. The effect of noise on the input-output relationship of a neuron. (A) The standard threshold-linear relationship between membrane potential and spike rate observed in most cells. (B) In the presence of noise the curve is effectively shifted left or right on different stimulus trials. That is, when noise depolarizes the cell, a stimulus can be that much smaller and still evoke action potentials. When the noise is evenly distributed around the resting potential, then the average input-output curve over many trials can be approximated by a power law (thick curve).



There is, however, a physiological reason why a flat threshold is not an accurate picture of the relationship between membrane voltage and action potential generation. This is simply the trial-to-trial variability of the responses of neurons (Anderson, Carandini, et al., 2000). The tuning curves of figure 21.2 are constructed from the average responses to many repeats of the same stimulus. For example, the spike threshold is often near or above the largest average response to a stimulus at the optimal orientation and the highest contrast. But such a stimulus, and many less effective ones as well, can evoke spikes, because on some individual trials the membrane potential exceeds threshold. The major difference between a high-contrast and low-contrast stimulus is not so much the size of the responses elicited but on how many trials a suprathreshold response occurs. The effect of this variability in response is shown in figure 21.3. A single threshold value can be represented as a linear function with a non-zero x-intercept (figure 21.3A). When the noise has depolarized the cell, it is as if the threshold were shifted down slightly for that trial; that is, the stimulus to depolarize the membrane potential less in order to evoke spikes. When the noise hyperpolarizes the cell, the opposite is true. The average input-output function of the cell is derived by averaging together the threshold functions of all trials (figure 21.3B). This average is no longer a standard linear function but rather can be well approximated by a power law (Hansel and van Vreeswijk, 2002; Troyer, 2002). This smoothed curve, when added to the Gaussian-like orientation tuning curves at different contrasts, narrows each one equally and thereby maintains contrast invariance both in membrane voltage and in action potential rate. The power law input-output relation and the Gaussian-shaped tuning curves are the only curves that show this property.

Voltage-to-action potential transformation

The importance of the action potential threshold in converting membrane potential responses into the output of cortical cells has been highlighted already in this chapter. As mentioned earlier, an important consequence of antiphase inhibition, supported by the data, is that the orientation tuning of the synaptic input to cortical neurons should be contrast invariant. However, the orientation tuning of extracellularly measured action potentials is also contrast invariant. The application of a single threshold to the contrast-dependent tuning curves in figure 21.2C will result in an iceberg effect that appears to broaden orientation tuning with increasing contrast. It is unlikely that the action potential threshold of neurons changes in response to stimuli of different contrasts.

The lack of a depolarizing response at high contrasts and nonpreferred orientations.

gratings preceded by cortical stimulation are reduced in amplitude, both because the shock evokes a shunt in the recorded cell and because the components of the response mediated by other cortical cells are suppressed by the shock. The relative size of the responses to different orientations, however, is largely unaffected, so that orientation selectivity is not much changed. Figure 21.6*B* and *D* show three tuning curves; one taken from drifting gratings at 12 orientations, one taken from the responses to flashed gratings of *A* and *C* recorded without cortical shock, and one taken from the responses to flashed gratings with cortical shock. The tuning curves all have comparable widths after being normalized responses to flashed gratings with cortical shock. A comparison of the width of to their peak amplitudes. A comparison of the width of tuning with and without cortical shock for nine cells is shown in figure 21.6*E*. Since orientation tuning of synaptic potentials was found to be unchanged during the suppression of cortical activity, it seems likely that the aggregate input from geniculate relay cells is by itself as well tuned for orientation as the total input from cortex and geniculate combined. These results strongly support a dominant role for feedforward input in determining orientation tuning of simple cells. Although the cortical circuit clearly has role in amplifying visual responses, these experiments show that orientation selectivity can be maintained even when the cortical network is disrupted.

average of 20 stimulus cycles. (*D*) Same as *C* but with the cortex cooled. Note the difference in vertical scale from *C*. (*E*) Orientation tuning curves drawn from the traces of *C* and *E*. Each point is the peak-to-peak amplitude of the first harmonic of the Fourier transform of the corresponding record. Cooling was done by a Reliance device attached to a copper plate, the end of which (4 × 4 mm) was placed on the surface of the brain. Intracellular recording electrode were inserted into the cortex through a small hole in the plate.

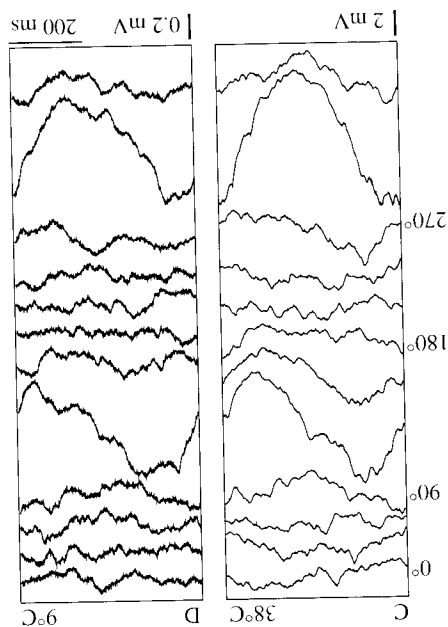
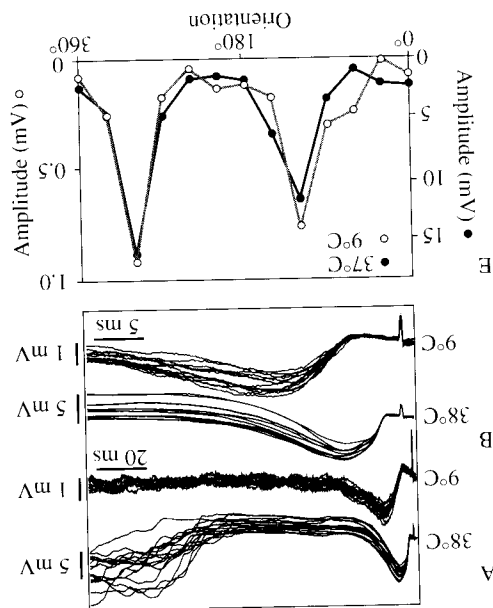


FIGURE 21.5. (*A*) The responses of a cortical simple cell to electrical stimulation of the LGN with the cortex at normal and cooled temperatures. During cooling, the initial evoked EPSP is reduced in size by fivefold and increased in latency. The long-lasting inhibitory rebound excitation, (*B*) The traces in *A* at a higher horizontal gain. (*C*) The response of the cell to drifting sinusoidal gratings at different orientations. Each response shown is the



in a second experiment (Chung and Ferster, 1998), electrical stimulation rather than cooling was used to inhibit cortical activity. The stimulation induces a strong, long-lasting which visual stimulation fails to evoke cortical spikes. Inputs recorded during the inhibition should therefore not from cortical cells but almost purely from geniculate input. The responses of two cortical simple cells to flashed gratings of different orientation are shown in figure 21.5*A* and *C*, with and without a preceding shock in the cortex nearby (thick and thin traces). The responses to the

responses are reduced 10-fold in amplitude by the cooling to compare the vertical scale bars in *B* and *C*, and yet the orientation tuning is hardly changed. That the shape of the orientation tuning has hardly changed can be seen qualitatively from the tuning curves drawn from the two sets of traces (figure 21.5*E*). These results were typical for 10 cells studied. They suggest that about 30% of the input to simple cells arises from the LGN, with the other 70% of the input coming from other cortical neurons. But there was little evidence that this cortical input was required to make cortical cells orientation selective, since the geniculate input remaining during cooling is fully orientation selective.

Another experimentally testable difference between forward and feedback models of orientation tuning concerns the time course of orientation-tuned responses to brief stimuli. Feedback models, which propose that initial excitatory input to the cortex is largely untuned, also predict that as the initial responses are propagated through the cortical circuitry and fed back as orientation-selective signals to the cells, the orientation tuning of the response will be seen to sharpen. Such an effect has been reported in extracellular recordings in monkey visual cortex (Ringach et al., 1997; Shapley et al., 2003). Although most cells in the input layer 4β were not found to sharpen, some cells in other layers showed a change in their tuning characteristics over time, consistent with feedback models. Another extracellular study (Mazer et al., 2002) failed to find a change in the shape of orientation tuning over time, but the time sampling of their data may have been too coarse to resolve such changes. A study performed intracellularly in the cat (Gillespie et al., 2001) found no sharpening of orientation tuning over time.

Dynamics of orientation tuning

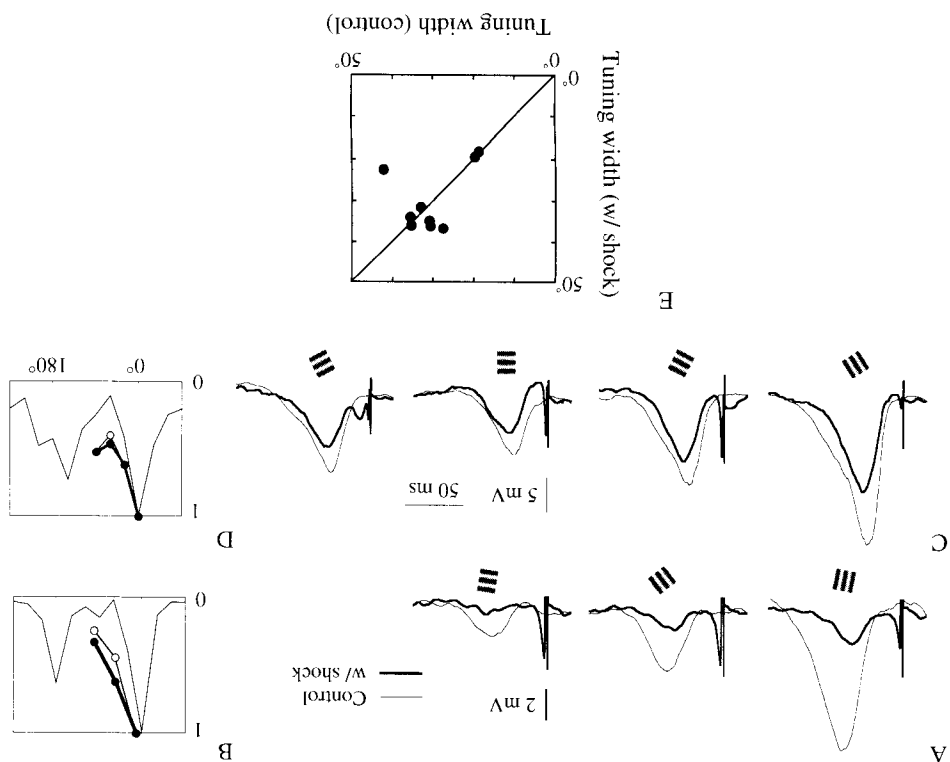
What remains is therefore likely to arise almost exclusively from cell and by suppressing the spike response of other cortical cells. amplitude of the response by introducing a shunt into the recorded cortex was delivered at the end of the flash. The shock reduces the control data. For the thick traces, an electrical stimulus to nearby orientation is indicated below each set of traces. Thin traces are flashed gratings confined to the classic receptive field. Grating derived from a simple cell to briefly (20 ms)

Although there is not universal agreement on this point, there is a great deal of evidence that orientation selectivity in simple cells of the cat visual cortex is determined largely by the spatial organization of geniculate input, just as Hubel

Conclusions

stimuli that were used. contrast responses to the sudden onset of the flashed bar (Pei et al., 1994) were complicated by orientation-nonspecific course of orientation selectivity of the membrane potential contrast changes. Earlier attempts to measure the time methods in which orientation was changed without large altering the selectivity of the synaptic inputs. These recent tuning of the extracellularly measured response without the action potential threshold will narrow the orientation discussed earlier, shifting the membrane potential relative to sample showed a subtractive reduction in membrane potential over time, consistent with an increase in inhibition. A in cells of any layer. However, about 50% of cells in the

synaptic excitation from the LGN. (B) Orientation tuning curves derived from the responses in A (open and closed circles) and from drifting gratings at 12 different orientations. Each of the three curves is normalized to its own peak. (C and D) Similar data to and B for a second simple cell. (E) A comparison of the width tuning for control and shock data for nine cells. To obtain the Gaussian.



- DeAngelis, G. C., L. Chazawa, et al., 1993. Spatiotemporal organization of simple-cell receptive fields in the cat's striate cortex. II. Linearity of temporal and spatial summation. *J. Neurophysiol.* 69:1118-1135.
- Eysel, U. T., 1992. Lateral inhibitory interactions in areas 17 and 18 of the cat visual cortex. *Pog. Brain Res.* 90:407-422.
- Eysel, U. T., J. M. Crook, et al., 1990. GABA-induced remote inactivation reveals cross-orientation inhibition in the cat striate cortex. *Exp. Brain Res.* 80:626-630.
- Eysel, U. T., I. A. Shevlevy, et al., 1998. Orientation tuning and receptive field structure in cat striate neurons during local blockade of intracortical inhibition. *Neuroscience* 84:25-36.
- Festerer, D., 1986. Orientation selectivity of synaptic potentials in neurons of cat primary visual cortex. *J. Neurosci.* 6:1284-1301.
- Festerer, D., 1988. Spatially opponent excitation and inhibition in simple cells of the cat visual cortex. *J. Neurosci.* 8:1172-1180.
- Festerer, D., S. Cheng, and H. Wierwyl, 1996. Orientation selectivity of thalamic input to simple cells of cat visual cortex. *Nature* 380:249-252.
- Festerer, D., and S. Lindström, 1983. An intracellular analysis of geniculocortical connectivity in area 17 of the cat. *J. Physiol.* 342:181-215.
- Friedl, T. F., K. A. Maritz, et al., 1989. Arborisation pattern and postsynaptic targets of physiologically identified thalamocortical afferents in striate cortex of the macaque monkey. *J. Comp. Neurol.* 289:313-336.
- Gardner, J. L., A. Azari, et al., 1999. Linear and nonlinear contributions to orientation tuning of simple cells in the cat's striate cortex. *Vis. Neurosci.* 16:1113-1121.
- Griespe, D. C., I. Lampl, et al., 2001. Dynamics of the orientation-tuning-membrane potential response in cat primary visual cortex. *Vis. Neurosci.* 18:1014-1019.
- Hammox, P., and C. J. Povrett, 1990. Influence of spatial frequency on tuning and bias for orientation and direction in the cat's striate cortex. *Vision Res.* 30:3359-369.
- Hansel, D., and C. van Vreeswijk, 2002. How noise contributes to contrast invariance of orientation tuning in cat visual cortex. *J. Neurosci.* 22:5118-5128.
- Hirsch, J. A., J. M. Aloosio, et al., 1998. Synaptic integration in striate cortical simple cells. *J. Neurosci.* 18:9517-9528.
- Hübner, D. H., and T. N. Wiesel, 1962. Receptive fields, binocular interaction and functional architecture in the cat's visual cortex. *J. Physiol. (Lond.)* 160:106-134.
- Hübner, D. H., and T. N. Wiesel, 1972. Laminar and columnar distribution of geniculocortical fibers in the macaque monkey. *J. Comp. Neurol.* 146:421-450.
- Jones, J. P., and L. A. Palmer, 1987. The two-dimensional spatial structure of simple receptive fields in cat striate cortex. *J. Neurophysiol.* 58:1187-1211.
- Lampl, I., J. S. Anderson, et al., 2001. Prediction of orientation selectivity from receptive field architecture in simple cells of cat visual cortex. *Neuron* 30:263-274.
- Mazer, J. A., W. E. Skaggs, et al., 2002. Spatial frequency and orientation tuning dynamics in area V1. *Proc. Natl. Acad. Sci. USA* 99:1645-1650.
- McLaughlin, D., R. Shapley, et al., 2000. A neuronal network model of macaque primary visual cortex (V1): Orientation selectivity and dynamics in the input layer 4Calpha. *Proc. Natl. Acad. Sci. USA* 97:8087-8092.
- Mittler, K. D., and T. W. Thorey, 2002. Neural noise can explain expansive, power-law nonlinearity in neural response functions. *J. Neurophysiol.* 87:653-659.
- Mooser, H., J. E. Mooser, et al., 2003. Emergent properties of layer 4 neurons reflect the collinear arrangement of horizontal connections in the streptococcus visual cortex. *J. Neurosci.* 23:2947-2960.
- Ng, S., and D. Festerer, 1998. Strength and orientation tuning of the thalamic input to simple cells revealed by electrically evoked suppression. *Neuron* 20:1177-1189.
- Ogawa, E. M., 1998. Local circuits in primary visual cortex of the macaque monkey. *Annu. Rev. Neurosci.* 21:47-74.
- Rakshit, M., and D. Festerer, 2000. Membrane potential and firing rate in cat primary visual cortex. *J. Neurosci.* 20:470-484.
- Schiffman, H. J., E. Mooser, et al., 2003. Emergent properties of layer 4 neurons reflect the collinear arrangement of horizontal connections in the streptococcus visual cortex. *J. Neurosci.* 23:2947-2960.
- Treisman, A., and D. Festerer, 1995. Theory of orientation tuning in visual cortex. *Proc. Natl. Acad. Sci. USA* 92:3844-3848.
- Van der Horst, E. M., 1998. Local circuits in primary visual cortex of the macaque monkey. *Annu. Rev. Neurosci.* 21:47-74.
- Wiesel, T. N., and D. Hubner, 1962. Receptive fields, binocular interaction and functional architecture in the cat's visual cortex. *J. Physiol. (Lond.)* 160:106-134.
- Wiesel, T. N., and D. Hubner, 1972. Laminar and columnar distribution of geniculocortical fibers in the macaque monkey. *J. Comp. Neurol.* 146:421-450.
- Xiao, J. S., L. Lampl, et al., 2000. The contribution of noise to contrast invariance of orientation tuning in cat visual cortex. *J. Neurophysiol.* 84:909-926.
- Xiao, J. S., M. Carandini, et al., 2000. Orientation tuning of output conductance, excitation, and inhibition in cat primary visual cortex. *J. Neurophysiol.* 84:909-926.
- Xiao, J. S., M. Carandini, et al., 2000. Orientation tuning of modular visual neurons in the cat. *Exp. Brain Res.* 101:413-426.
- Xiao, J. S., and A. B. Bonds, 1994. Inactivation of the infrastriate cortex broadens orientation tuning of supra-striate visual neurons in the cat. *Exp. Brain Res.* 101:413-426.

- MONIER, C., F. CHAVANE, et al., 2003. Orientation and direction selectivity of synaptic inputs in visual cortical neurons: a diversity of combinations produces spike timing. *Neuron* 37:663-680.
- MOVSHON, J. A., I. D. THOMPSON, et al., 1978. Spatial summation in the receptive fields of simple cells in the cat's striate cortex. *J. Physiol.* 283:53-77.
- NELSON, S., L. TOTH, B. SHETH, and M. SUR, 1994. Orientation selectivity of cortical neurons during intracellular blockade of inhibition. *Science* 265:774-777.
- PEI, X., T. R. VIDYASAGAR, M. VOLGUSHEV, and O. GREUTZFELDT, 1994. Receptive field analysis and orientation selectivity of post-synaptic potentials of simple cells in cat visual cortex. *J. Neurosci.* 14:7130-7140.
- PRITZGER, B., and A. B. BONDS, 1995. Dynamic differentiation of GABA_A-sensitive influences on orientation selectivity of complex cells in the cat striate cortex. *Exp. Brain Res.* 104:81-88.
- REID, R. C., and J. M. ALONSO, 1995. Specificity of monosynaptic connections from thalamus to visual cortex. *Nature* 378:281-284.
- RINGACH, D. L., M. J. HAWKES, and R. SHAPLEY, 1997. Dynamics of orientation tuning in macaque primary visual cortex. *Nature* 387:281-284.
- RINGACH, D. L., R. M. SHAPLEY, et al., 2002. Orientation selectivity in macaque V1: Diversity and laminar dependence. *J. Neurosci.* 22:5639-5651.
- SATO, H., N. KATSUYAMA, et al., 1996. Mechanisms underlying orientation selectivity of neurons in the primary visual cortex of the macaque. *J. Physiol.* 494:757-771.
- SOLAR, G., and R. D. FREEMAN, 1982. Orientation selectivity in the cat's striate cortex is invariant with stimulus contrast. *Exp. Brain Res.* 46:457-461.
- SHAPLEY, R., M. HAWKES, et al., 2003. Dynamics of orientation selectivity in the primary visual cortex and the importance of cortical inhibition. *Neuron* 38:689-699.
- SILVER, A. M., 1975. The contribution of inhibitory mechanisms to the receptive field properties of neurons in the striate cortex of the cat. *J. Physiol.* 250:303-329.
- SOMERS, D. C., S. B. NELSON, and M. SUR, 1995. An emergent model of orientation selectivity in cat visual cortical simple cells. *J. Neurosci.* 15:5448-5465.
- TOLHURST, D. J., and A. F. DEAN, 1987. Spatial summation by simple cells in the striate cortex of the cat. *Exp. Brain Res.* 66:607-620.
- TOLHURST, D. J., and A. F. DEAN, 1990. The effects of contrast on the linearity of spatial summation of simple cells in the cat's striate cortex. *Exp. Brain Res.* 79:582-588.
- TROYER, T. W., A. E. KUKOROWSKI, et al., 1998. Contrast-invariant orientation tuning in cat visual cortex: Thalamocortical input tuning and correlation-based intracortical connectivity. *J. Neurosci.* 18:5908-5927.
- USREY, W. M., M. P. SCERNIAK, et al., 2003. Receptive fields and response properties of neurons in layer 4 of ferret visual cortex. *J. Neurophysiol.* 89:1003-1015.
- VOLGUSHEV, M., T. R. VIDYASAGAR, et al., 1996. A linear model fails to predict orientation selectivity of cells in the cat visual cortex. *J. Physiol.* 496(Pt 3):597-606.
- WEBSTER, M. A., and R. L. DE VALOIS, 1985. Relationship between spatial-frequency and orientation tuning of striate-cortex cells. *J. Opt. Soc. Am.* 4:1124-1132.